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| Step 4/6  Magnetic Immunoprecipitation  Time  30m +2h/Overnight  Vol.  100µl/IP | 4 Magnetic Immunoprecipitation  1) Briefly spin the tubes/8-strips containing the antibody-coated beads to bring down liquid caught inside the lid.  2) Place tubes in the ice-cold Magnetic Rack (cooled by placing on ice or in refrigerator), wait for 1 minute.  3) Discard the supernatant. Keep the pellet of antibody-coated beads.  4) Add 100 µl of diluted sheared chromatin (from step2 or step 3) to each IP tube.  5) Keep 10 µl of diluted chromatin as Input sample at 4°C (or stored at -20ºC).  !! Attention: Avoid bubbles. It’s very easy to pipette bubbles as the Buffer contains detergent. Use reverse pipetting technique to reduce the loss of material. Measure the volume before adding the chromatin, always take Input sample first, then IPs (IgG last)  6) Invert the tubes several times make sure beads are re-suspended.  7) Incubate samples at 4ºC under constant rotation on a rotator at 40 rpm for 2 hours up to overnight.  Pause Point: Samples could be incubated at 4 ºC overnight. Longer incubation may increase background. |